

Enhancement of Antiviral CD8⁺ T-Cell Responses and Complete Remission of Metastatic Melanoma in an HIV-1-Infected Subject Treated with Pembrolizumab

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Abstract: Background: Pembrolizumab is an immune checkpoint inhibitor against programmed cell death protein-1 (PD-1) approved for therapy in metastatic melanoma. PD-1 expression is associated with a diminished functionality in HIV-1 specific-CD8⁺ T cells. It is thought that PD-1 blockade could contribute to reinvigorate antiviral immunity and reduce the HIV-1 reservoir. Methods: Upon metastatic melanoma diagnosis, an HIV-1-infected individual on stable suppressive antiretroviral regimen was treated with pembrolizumab. A PET-CT was performed before and one year after pembrolizumab initiation. We monitored changes in the immunophenotype and HIV-1 specific-CD8⁺ T-cell responses during 36 weeks of treatment. Furthermore, we assessed changes in the viral reservoir by total HIV-1 DNA, cell-associated HIV-1 RNA, and ultrasensitive plasma viral load. Results: Complete metabolic response was achieved after pembrolizumab treatment of metastatic melanoma. Activated CD8⁺ T-cells expressing HLA-DR⁺/CD38⁺ transiently increased over the first nine weeks of treatment. Concomitantly, there was an augmented response of HIV-1 specific-CD8⁺ T cells with TNF production and poly-functionality, transitioning from TNF to an IL-2 profile. Furthermore, a transient reduction of 24% and 32% in total HIV-1 DNA was observed at weeks 3 and 27, respectively, without changes in other markers of viral persistence. Conclusions: These data demonstrate that pembrolizumab may enhance the HIV-1 specific-CD8⁺ T-cell response, marginally affecting the HIV-1 reservoir. A transient increase of CD8⁺ T-cell activation, TNF production, and poly-functionality resulted from PD-1 blockade. However, the lack of sustained changes in the viral reservoir suggests that viral reactivation is needed concomitantly with HIV-1-specific immune enhancement.

Keywords: Immune checkpoint inhibitors; pembrolizumab; HIV-1 reservoir; HIV-specific CD8⁺ T cells; HIV-1 curative strategies

1. Introduction

Inhibitory receptors (iRs) or immune checkpoint molecules play a key role in the regulation of persistent immune activation during cancer and chronic infections to avoid self-damage. Immune checkpoint inhibitors (ICIs) are monoclonal antibodies that target iRs to reverse T-cell exhaustion due to continuous antigen stimulation. Pembrolizumab (Keytruda[®], MSD) is an ICI directed against programmed cell death protein-1 (PD-1), which blocks the interaction with its ligands PD-L1 and PD-L2. These ligands are overexpressed on activated antigen presenting cells as well as cancer cells, and their blockade promotes T-cell activation against tumor cells [1]. Currently, pembrolizumab, approved by FDA, has become a first-line treatment against metastatic or unresectable melanoma providing a five-year survival rate between 34% and 41% [2].

Expression of PD-1 has also been associated with dysfunctional HIV-1 specific-CD8⁺ T-cell responses and disease progression. In this context, in vitro PD-1 blockade has demonstrated a recovery of HIV-1 specific-CD8⁺ T-cell functionality [3,4]. On the other hand, iR-expressing CD4⁺ T cells have been associated with cell-based measures of viral persistence in HIV-1-infected patients on antiretroviral therapy (ART) [5,6], and ex vivo experiments have shown an enhancement of viral production by CD4⁺ T cells after PD-1 blockade [7,8]. Therefore, ICIs could impact on the HIV-1 reservoir through the so-called 'shock and kill' mechanism; i.e., reactivating latent HIV-1 provirus from infected CD4⁺ T cells and reversing exhausted HIV-1 specific-CD8⁺ T cells against HIV-1 producing cells, altogether resulting in a potential reduction of the reservoir that ultimately might help viral remission [9].

Different case reports, and a couple of case series, suggest that ICI therapy appears to be safe and efficacious in HIV-1-infected individuals with advanced stage cancer [10], although ongoing prospective trials of ICI need to confirm these findings [11–13]. However, the effectiveness of ICI therapy to help eliminate the viral reservoir in ART-treated individuals is still controversial and only based on limited case reports. These cases range from showing no major changes in either HIV-1 specific-CD8⁺ T-cell responses or HIV-1 persistence [14], transient enhancement of HIV-1 specific-CD8⁺ T cells with no variation in viral persistence [15,16], transient increase in viral transcription without changes in viral reservoirs [17], or one case of depletion of the HIV-1 reservoir [18].

Here, we report a case of an HIV-1-infected individual on ART that received pembrolizumab treatment for metastatic melanoma. We combine the description of clinical response to melanoma after pembrolizumab treatment with a detailed characterization of functional T-cell responses as well as the therapeutic impact on viral persistence.

Case Report

A 46-year-old man was diagnosed with HIV-1 infection in 2008, immediately receiving suppressive ART (Figure 1A). In 2011, his ART was simplified to ritonavir-boosted darunavir. In June 2016 he was diagnosed with an amelanotic melanoma on the right hemithorax, with palpable right axillary lymphadenopathy. Staging positron emission tomography-computed tomography (PET-CT) and neurologic magnetic resonance image (MRI) showed no evidence of distant metastatic disease. In September 2016, an axillary lymphadenectomy was performed with involvement of 3/22 lymph nodes by melanoma. There were no mutations in the codon V600 of the BRAF gene, NRAS in exon 2 or 3, or in the KIT gene (PCR Sanger sequencing, Therascreen BRAF Pyro Kit, Qiagen; Hilden, Germany). Repeated PET-CT showed disease progression with hypermetabolic focal lesions in the right axillary, right pleura, and D4 vertebral body (Figure 1B1–B3). In October 2016, the subject initiated pembrolizumab (2 mg/kg every three weeks). At the onset of melanoma, the ART was switched to raltegravir 400 mg twice daily and tenofovir fumarate/emtricitabine to avoid drug interactions.

copies/mL). The subject had never presented any AIDS-associated illness. Genotyping of HLA class I loci indicated homozygosity for HLA-A*02, HLA-B*07, and HLA-C*07. HLA class II loci showed

A new PET-CT after 12 cycles of pembrolizumab treatment showed complete disappearance of the right axillary, right pleura, and D4 vertebral body lesions (Figure 1B4–B5). The infusions were His CD4⁺ T-cell count tolerated, was without requiring systemic steroids. Plasma for HIV-1 RNA inflammation was suppressed to pembrolizumab (<40 copies/mL). Peripheral blood mononuclear cells (PBMCs) and plasma samples were collected before each pembrolizumab administration at weeks (w) 0, 3, 9, 18, 27, and 36 (Figure 1A). Additionally, we evaluated the short-term effect of pembrolizumab in a sample one day after pembrolizumab HLA-DRB1*15, administration and HLA-DQB1*06 at w18(w18+1). The subject provided informed consent and the institutional review board approved the investigational protocol (ref PI-18-229).

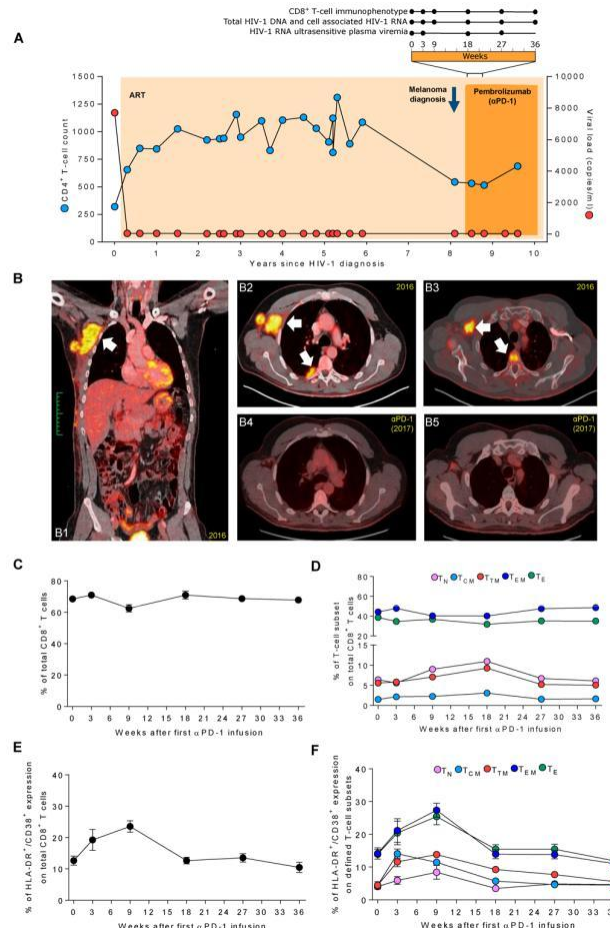


Figure 1. Clinical follow-up and impact of pembrolizumab administration in an HIV-1-infected individual on longitudinal active antiretroviral treatment (ART) who developed metastatic melanoma; (A) individual on longitudinal active antiretroviral treatment (ART) (blue circles) who and HIV developed viral load (red circles) melanoma;

(A) Longitudinal analysis in years of CD4⁺ T-cell count (blue circles) and HIV-1 viral load (red circles) since HIV-1 diagnosis. Light orange area indicates the time on ART and dark orange area depicts the time of pembrolizumab administration under ART. Blue arrow indicates melanoma diagnosis. Top bar represents the cycles of pembrolizumab administrations and the biological samples that were analyzed at 0, 3, 9, 18, 27, and 36 weeks. Black lines indicate measures in CD8⁺ T-cell immunophenotype, total HIV-1 DNA, cell-associated (CA) HIV-1 RNA and, ultrasensitive viral load in plasma (HIV-1 RNA (usVL)); (B) coronal (left panel) and axial (right panel) images of positron emission tomography with 2-deoxy-2-(fluorine-18) fluoro-d-glucose integrated with computed tomography (18F-FDG PET/CT). B1–B3 images depict the lesions before pembrolizumab administration (27 September 2016) by the increased uptake of 18F-FDG in the right axillary lymphoid node and local soft tissue invasion in pleura and osteoblastic spine metastasis. B4 and B5 panels show serial imaging after one year of pembrolizumab administration (27 September 2017) of axial baseline and indicating sustained complete metabolic response and the resolution of the lesions. For B1 to B5, white arrows indicate the localization of the metastatic lesions of melanoma; (C) changes in total CD8⁺ T cells after the first pembrolizumab administration in weeks; (D) changes in CD8⁺ T-cell subsets; (E) analysis of total CD8⁺ T cells expressing HLA-DR⁺/CD38⁺; and (F) CD8⁺ T-cell subsets expressing HLA-DR⁺/CD38⁺. TN, naïve; TCM, central memory; TTM, transitional memory; TEM, e effector memory; TE, e effector. Results are expressed as the mean of two measurements. RAL, Raltegravir; TDF/FTC, truvada.

A new PET-CT after 12 cycles of pembrolizumab treatment showed complete disappearance of the right axillary, right pleura, and D4 vertebral body lesions (Figure 1B4–B5). The infusions were well tolerated, without requiring systemic steroids for signs of local inflammation to pembrolizumab.

Peripheral blood mononuclear cells (PBMCs) and plasma samples were collected before each pembrolizumab administration at weeks (w) 0, 3, 9, 18, 27, and 36 (Figure 1A). Additionally, we evaluated the short-term effect of pembrolizumab in a sample one day after pembrolizumab administration at w18 (w18+1). The subject provided informed consent and the institutional review board approved the investigational protocol (ref PI-18-229).

2. Experimental Section

2.1. Immune Phenotype of Total CD8⁺ and HIV-1 Specific-CD8⁺ T Cells

To monitor changes in CD8⁺ T cells and HIV-1 specific-CD8⁺ T-cell responses after pembrolizumab initiation, PBMCs were stimulated with HIV-1 Gag peptide pool (2 g/peptide/mL, EzBiolab, Carmel, IN, USA), Staphylococcal enterotoxin B (SEB) (1 g/mL, Sigma-Aldrich, Madrid, Spain), or no stimuli, in the presence of CD28/49d co-stimulatory molecules (1 g/mL, BD, Madrid, Spain), Monensin A (1 g/mL, BD), and CD107a (PE-Cy5, clone eBioH4A3, BD) for 6 h at 37 °C in a 5% CO₂ incubator and rested overnight at 4 °C. After incubation, cells were washed with PBS 1X and stained with a viability dye (APC-Cy7, Thermo Fisher Scientific) for 30 min at room temperature (RT). Cells were surface stained for 30 min at RT with anti-human antibodies for CD3 (A700, clone UCHT1, BD), CD4 (APC-Cy7, clone SK3, BD), CD8 (V500, clone RPA-T8, BD), CD45RA (BV786, clone HI100, BD), CCR7 (PE-CF594, clone 150503, BD), CD27 (BV605, clone L128, BD), PD-1 (BV421, clone EH12.1, BD), CD38 (PE, clone HB-7, BD), and HLA-DR (A647, clone L243, BioLegend, London, UK). Afterwards, cells were fixed with Fix/Perm Buffer A (Thermo Fisher Scientific) for 15 min at RT and intracellular stained with Fix/Perm Buffer B and antibodies for TNF (PE-Cy7, clone MAb11, BioLegend), IFN (BV711, clone B27, BD), and IL-2 (BV650, clone MQ1-17H12, BD) for 20 min at RT. Finally, cells were resuspended and fixed in formaldehyde 1% and acquired on LSR Fortessa cytometer using FACSDiVa software (BD). Data were analyzed with FlowJo software v10. We excluded dump and CD4⁺ T cells, and gates were defined using fluorescence minus one (FMO) controls. Surface markers were measured in total CD8⁺ T cells and CD8⁺ T-cell subsets including naïve (TN), central memory (TCM), transitional memory (TTM), effector memory (TEM), and effector (TE), as previously described [19]. The frequency of each surface marker was expressed as the mean of two independent replicates. For HIV-Gag specific-CD8⁺ T-cell cytokine production, only positive replicates were included. GraphPad Prism v4.0 software was used for graph plotting. Pestle and Spice software [20] was applied to visualize poly-functionality of HIV-1 specific-CD8⁺ T-cell responses for the evaluation of cytokine pattern distribution.

2.2. Total HIV-1 DNA, Cell-Associated HIV-1 RNA, and Ultrasensitive Viral Load

To evaluate the size of the proviral reservoir, total HIV-1 DNA was measured in purified peripheral CD4⁺ T-cell lysates using droplet digital PCR (ddPCR), as previously described [21]. Briefly, 5⁰ long terminal repeat (5⁰LTR) region was amplified in duplicate, and the RPP30 housekeeping gene was quantified in parallel to normalize sample input. Raw ddPCR data were analyzed using the QX100 Droplet Reader and the QuantaSoft v.1.6 software (Bio-Rad, Hercules, CA, USA).

Also, viral transcription was evaluated by quantification of cell-associated HIV-1 RNA (ca-HIV-1 RNA) in purified CD4⁺ T cells, using one-step reverse-transcription ddPCR22. The 5⁰-LTR gene and the housekeeping gene TATA-binding protein (TBP) were measured in parallel. Residual low-level viremia was determined through ultrasensitive viral load (usVL) by ultracentrifugation of 5–6 mL of plasma and quantified using the Abbott Real-Time HIV-1 assay (Abbott Molecular Inc., Des Plaines, IL, USA), as previously described [22,23]. All determinations were done on equivalent number of CD4⁺ T cells.

3. Results

3.1. Pembrolizumab Administration Induces a Marked and Transient Increase of HLA-DR⁺/CD38⁺ Expression in TEM and TE CD8⁺ T Cells

Immunotherapies are supposed to affect the distribution of immune cell populations and their activation status. We measured changes in the frequency of total CD8⁺ T cells (Figure 1C) and stable counts were observed despite a transient decrease at w9. The frequency of total CD8⁺ T cells was 68.5% at baseline and was contracted to 62.5% at w9, returning to baseline levels beyond w18. Regarding CD8⁺ T-cell subpopulations (Figure 1D), no major changes were observed in the distribution over the analyzed period. Only the expression of TN and TTM showed a transient increase between w9 and w18. When we evaluated the short-term effect at w18+1, we identified a slight decrease in total CD8⁺ T cells (70.9% to 67.4%) that was concomitant in TN, TCM, and TTM subpopulations. However, TE and TEM raised 7.3% and 7.2%, respectively, after 24 h of pembrolizumab infusion (Supplementary Materials Figure S1A).

Concerning the activation status, total CD8⁺ T cells showed increased expression of the activation markers HLA-DR and CD38 after the first pembrolizumab infusion [24,25] (Figure 1E). Activated CD8⁺ T cells raised progressively from w0 to w9 (12.7% to 23.6%) and returned to baseline levels at w18 (12.7%). This transient increase of HLA-DR⁺/CD38⁺ expressing cells was also observed in all the CD8⁺ T-cell subpopulations. The percentage of activated TN, TTM, TEM, and TE increased gradually after the first infusion and peaked at w9 (Figure 1F). Particularly, the expression of HLA-DR⁺/CD38⁺ in TEM and TE cells doubled from 14.2% to 27.4% and from 13.9% to 25.4%, respectively. Nevertheless, activated TCM cells peaked at w3 and their levels decreased progressively overtime. Regarding the short-term effect at w18+1, we observed a small increase in the activation status of total CD8⁺ T cells (12.7% to 13.9%) but no changes in CD8⁺ T-cell subsets (Supplementary Materials Figure S1B). Altogether, these data show that pembrolizumab transiently activates peripheral CD8⁺ T cells, more specifically effector cells, within the first few infusions in line with its immune therapeutic mechanism.

3.2. Enhancement of TNF Production in HIV-1 Specific-CD8⁺ T-Cell Responses after Pembrolizumab Initiation

In addition to the evaluation of changes in CD8⁺ T-cell populations and their activation status, we wanted to measure the magnitude and functional profile of HIV-1 specific responses. Thus, we performed a detailed characterization of HIV-1 specific-CD8⁺ T cells by a combination of surface lineage markers and intracellular cytokine detection in response to HIV-Gag. We found an increase in HIV-1 specific-CD8⁺ T-cells defined by an augment of TNF production between w0 and w9 (0.3% to 0.5%) after the first infusion of pembrolizumab (Figure 2A). The response decreased progressively to undetectable levels at w36. Interestingly, concomitant to TNF reduction, we observed a slight increase of IFN γ , CD107a, and IL-2 production by HIV-1 specific-CD8⁺ T cells at w18, which stayed that way until the end of the follow-up. Additionally, at w18+1 we observed an increase from 0.2% to 0.4% on TNF production by HIV-1 specific-CD8⁺ T cells. However, no changes were observed in CD107a, IFN γ , and IL-2 (Supplementary Materials Figure S1C). Also, we observed a high frequency of TNF, CD107a, IFN γ , and IL-2 cytokine-producing cells in response to CD8⁺ T-cell polyclonal activation with the SEB stimuli that transiently peaked at w3 (Figure 2B).

By using lineage expression markers, we tried to decipher the phenotype that underlined the peak of TNF production by HIV-1 specific-CD8⁺ T cells. We observed a decrease in CCR7, CD27, and an increase in HLA-DR and PD-1 markers between w0 and w9 (Figure 2C). These data indicate an effector profile of HIV-1 specific-CD8⁺ T cells responsible of TNF production at the peak of response to pembrolizumab.

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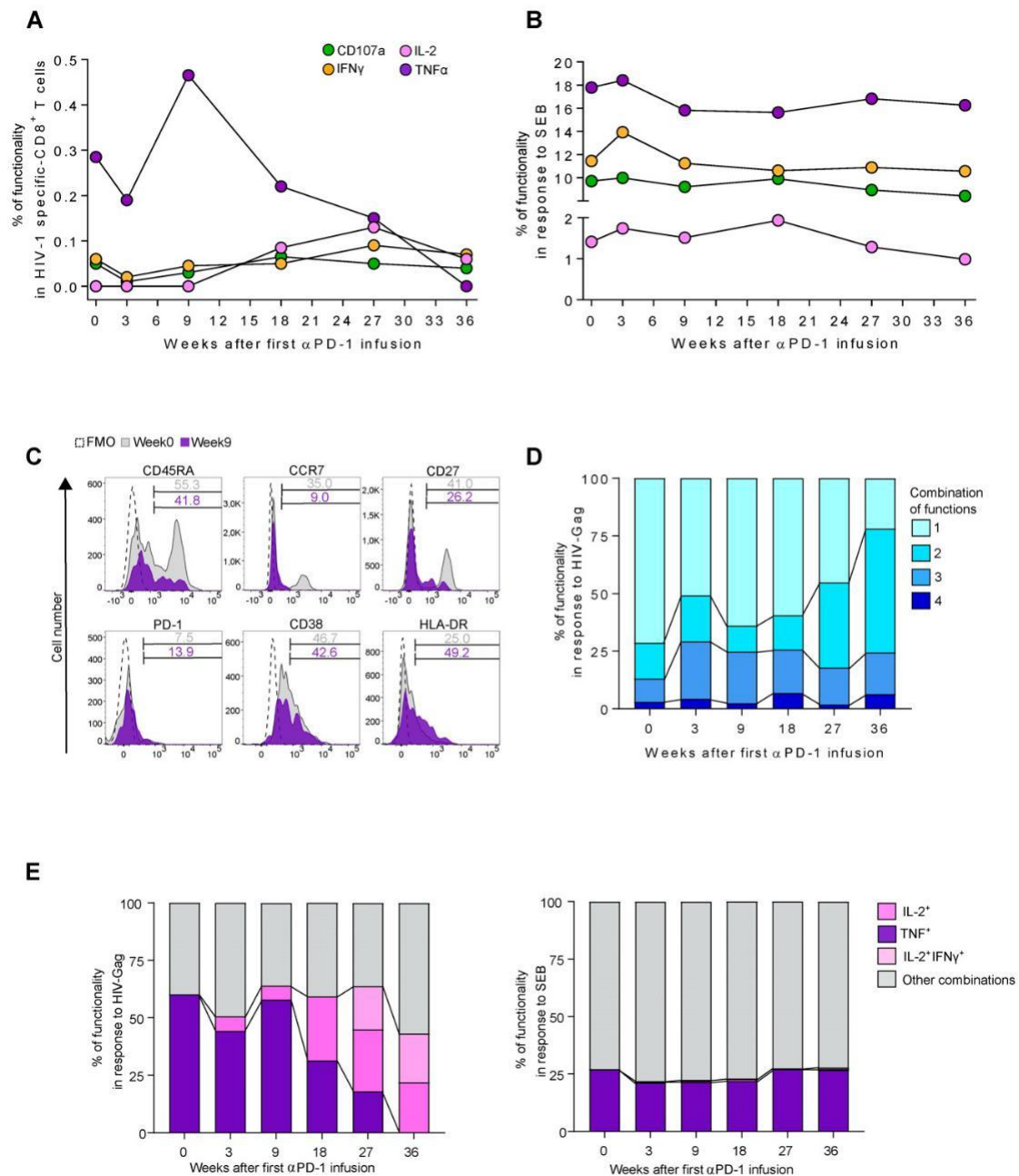


Figure 2. Immunophenotype of the HIV-1 specific-CD8⁺ T-cell response during pembrolizumab

Gag HIV-1 specific-CD8⁺ T-cells responses; (B) cytokine changes in response to Staphylococcal enterotoxin B (SEB); (C) histograms depicts the expression of CD45RA, CCR7, CD27, PD-1, CD38, and B (SEB); (C) histograms depicts the expression of CD45RA, CCR7, CD27, PD-1, CD38, and B (SEB); (D) poly-functionality expressed as the combination of one, two, three, or four cytokines in HIV-1 specific-CD8⁺ T cells; (E) detailed poly-functional profiles of HIV-1 specific-CD8⁺ T cells and under SEB stimulation. The bars in purple represent the evolution of TNF⁺ frequency overtime in single combination and pink represents the changes of IL-2⁺ and IL-2⁺IFN⁺ over the week 18.

3.3. Functional Switch from TNF to IL-2 Profile in HIV-1 Specific-CD8⁺ T-Cell Responses after Pembrolizumab Follow Up

By using a detailed functional profile of cytokine production based on TNF, IFN, CD107a surface expression, and IL-2 expression, we analyzed the evolution of HIV-1 Gag-specific CD8⁺ T-cell responses overtime (Figure 2D,E). Pembrolizumab raised the cell's polyfunctionality, expressed as any combination of two or more cytokines, from 28.7% at w0 to 78.2% at w36 (Figure 2D). Regarding combinations of three cytokines, although we observed peaks at w3 (24.9%) and w9 (22.3%), these combinations remained high until the end of the study. Indeed, detailed analyses showed a transient increase driven by HIV-1 specific-CD8⁺ T-cells expressing TNF (TNF⁺) during the first nine weeks of pembrolizumab treatment. Nonetheless, the TNF dominated profile switched to an IL-2 dominant profile (IL-2⁺ and IL-2⁺IFN⁺) since w9 (Figure 2E). In the short-term effect, we found an augment of TNF expressing cells (TNF⁺) at w18+1, concomitant with a reduction in IL-2⁺ production (Supplementary Materials Figure S1D,E). However, the combination IL-2⁺IFN⁺ seems to start to raise at this timepoint. These data contrast with those from polyclonal stimuli where no changes were observed. Thus, the initial release of TNF by HIV-1 specific-CD8⁺ T cells is switched to an IL-2 release profile in the long-term antiviral response.

3.4. Transient Decays of Total HIV-1 DNA

To monitor the perturbation in the HIV-1 CD4⁺ T-cell reservoir induced by pembrolizumab, we measured viral persistence by HIV-1 DNA, ca-HIV-1 RNA, and usVL. For all the analyzed samples, we detected total HIV-1 DNA with a median IQR of 1008 (862–1141) copies/million CD4⁺ T cells. We detected transient decays in total HIV-1 DNA from 1080 to 820 copies per million CD4⁺ T cells (24%) at w3, and 727 copies per million CD4⁺ T cells (32%) at w27 (Figure 3A). However, after each reduction, the levels of total HIV-1 DNA recovered to values similar to those of the baseline. Moreover, the expression of cell-associated unspliced HIV-1 RNA in CD4⁺ T cells was stable over time in all samples with a median level of 40 (36–47) ratio (HIV/TBP) x 1000 (Figure 3A).

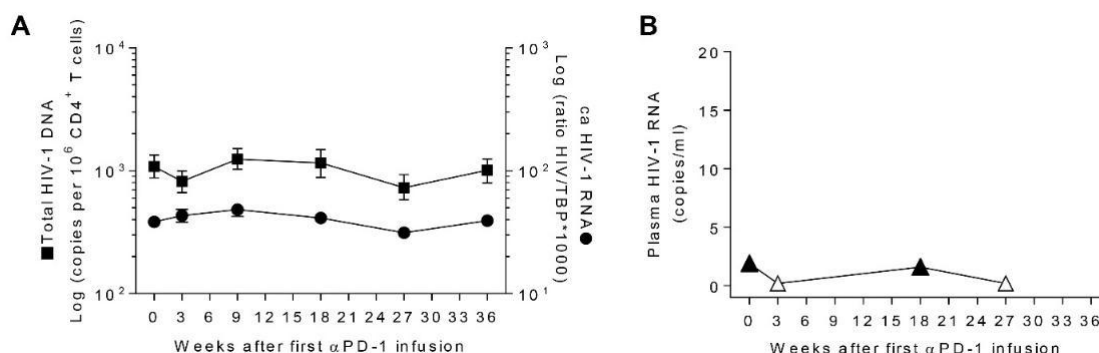


Figure 3. Longitudinal analysis of HIV-1 reservoir during pembrolizumab administration; (A) Total HIV-1 DNA (squares) and analysis of cell-associated HIV-1 RNA (circles) during pembrolizumab administration; measured by (A) ddPCR; Total

(B) ultrasensitive viral load in plasma (triangles). Open symbols represent determinations below the limit of quantification.

In order to determine the effect of pembrolizumab on HIV-1 reactivation, we measured HIV-1 RNA by

ultrasensitive viral load. We detected HIV-1 RNA in 50% of analyzed plasma samples. Interestingly, rather than seeing the production of new virions, we observed a reduction of viremia below the limit. Previous studies have hypothesized that ICIs might be possible candidates to the “shock and kill” curative strategy by the reactivation of the HIV-1 reservoir [7,8,26] and the concomitant to the transient reduction of total HIV-1 DNA observed. When we evaluated the short-term effect of enhancement of antiviral immune responses [3,4,27], leading to the clearance of reactivated cells and pembrolizumab at w18+1, we found levels of total HIV-1 DNA and ca-HIV-1 RNA within the same consequently reducing the HIV-1 reservoir size. However, the effect of PD-1 blockade in HIV-1 magnitude than at w18 (Supplementary Materials Figure S1F). Thus, we found transient decreases of infected individuals is incompletely understood, and the results are controversial, in part due to

limited experience in this clinical setting. Here, we not only show a case of complete remission of metastatic melanoma after treatment with pembrolizumab in a HIV-1-infected individual, but also an

enhancement of T-cell activation and the HIV-1 specific-CD8⁺ T-cell response. However, this T-cell invigoration was associated only with a transient perturbation on the HIV-1 reservoir size.

Pembrolizumab had a good safety profile and a sustained antitumoral response in this HIV-1-infected subject. Although the experience of pembrolizumab, as well as other ICIs, in people with

total proviral HIV-1 DNA and ultrasensitive plasma HIV-1 RNA at w3 and w27 after pembrolizumab treatment without a detectable cell-associated unspliced HIV-1 RNA in CD4⁺ T cells.

4. Discussion

Previous studies have hypothesized that ICIs might be possible candidates to the “shock and kill” curative strategy by the reactivation of the HIV-1 reservoir [7,8,26] and the concomitant enhancement of antiviral immune responses [3,4,27], leading to the clearance of reactivated cells and consequently reducing the HIV-1 reservoir size. However, the effect of PD-1 blockade in HIV-1-infected individuals is incompletely understood, and the results are controversial, in part due to limited experience in this clinical setting. Here, we not only show a case of complete remission of metastatic melanoma after treatment with pembrolizumab in a HIV-1-infected individual, but also an enhancement of T-cell activation and the HIV-1 specific-CD8⁺ T-cell response. However, this T-cell invigoration was associated only with a transient perturbation on the HIV-1 reservoir size.

Pembrolizumab had a good safety profile and a sustained antitumoral response in this HIV-1-infected subject. Although the experience of pembrolizumab, as well as other ICIs, in people with HIV-1 is limited, these results are in line with other preliminary studies assessing the safety of ICI therapies in HIV-1-infected individuals with advanced-stage cancer [10,11]. However, only a fraction of these patients with advanced melanoma benefit from ICI blockade [10,11], which highlights this case of complete remission of this metastatic melanoma. Immunologically, over the first nine weeks, we found an increase of HLA-DR⁺/CD38⁺ expressing cells, especially in TEM and TE CD8⁺ T-cell subsets, which had also been reported previously [18]. While previous studies only found IFN production by HIV-1 specific-CD8⁺ T cells [15,16,18], we describe for the first time the production of a peak of TNF production by HIV-1 specific-CD8⁺ T cells at w9 as a result of the PD-1/PD-L1 pathway disruption by pembrolizumab, in agreement with the initiation of an inflammatory response. TNF is a pleiotropic cytokine, mainly produced by activated cells, regulating the immune response, promoting inflammation, and inhibiting tumorigenesis [28].

After the peak of TNF, we observed a transition from TNF to an IL-2 cytokine profile consistent with the reduction of T-cell activation through the antigen receptor [29]. IL-2 plays a crucial role in the expansion of effector T cells that get activated upon antigen encounter, and regulates proliferation and homeostasis of T, B, and NK cell responses [28,30]. IL-2 helps to resolve the inflammatory response caused by pembrolizumab in the first weeks of treatment, contributing to the maintenance of HIV-1 specific-CD8⁺ T cells and generating a long-term memory response [30]. However, we cannot exclude an impact of pembrolizumab on other cell types including NK cells. Indeed, NK cell responses can have crucial immunotherapeutic effects upon PD-1/PD-L1 blockade in tumors with low levels of MHC-I expression [31]. Additional studies profiling functional immune cell responses during treatment with ICIs could bring new insights on the mechanism of these new therapies in the remission of cancer and viral diseases.

Despite observing two punctual drops on proviral DNA during the study, we were unable to detect a consistent decrease in the viral reservoir. This observation has been previously reported by other studies using ICIs in combination with ART [14–17]. These results differ from a single case report that shows a decrease from 369 to 30 copies HIV-1 DNA per million cells at day 120 after nivolumab administration [18]. Also, it has recently been reported that pembrolizumab led to transiently short-term small increase in unspliced cell-associated HIV-1 RNA in CD4⁺ T cells in vivo in individuals on ART [32]; however, due to differences in sampling times, we cannot corroborate such observations.

The absence of sustained reduction on proviral DNA in our study case might be due to the lack of expression of cell-associated HIV-1 RNA and the subsequent production of viral antigen needed for the targeting of infected cells by effector HIV-1 specific-CD8⁺ T cells [26]. Although there was an enhancement on HIV-1 specific-CD8⁺ T-cell responses, we did not observe a sustained reduction of viral reservoir in peripheral CD4⁺ T cells, suggesting that pembrolizumab was unable to effectively

reverse viral latency. Alternatively, any sustained reduction in the viral reservoir might have been unnoticed due to homeostatic proliferation of latently HIV-1-infected CD4⁺ T cells expressing PD-1, which might mask the detection of any effective “shock and kill” events under pembrolizumab treatment. Furthermore, the limitation of our analyses to peripheral blood samples underestimate the effects on lymphoid tissues where CD4⁺ T follicular helper cells expressing PD-1 play an important role as HIV-1 reservoirs [33]. Furthermore, data from ex vivo measurements on CD4⁺ T cells from ART-suppressed individuals suggest that pembrolizumab enhances the induction of HIV-1 production when combined with the latency reversing agent bryostatin, without providing data in reservoir clearance [8].

This study is restricted to peripheral blood samples from a single patient with complete control of viral replication under suppressive ART, which might have masked any beneficial effect of anti-PD1 treatment on plasma HIV viral load. These limitations should be carefully considered, in view of interindividual variability, especially in cancer patients. Ongoing clinical trials administrating ICI alone [32,34] or in combination with latency reversing agents to HIV-1-infected individuals on ART would help to clarify the potential contribution of ICIs in curative strategies.

Overall, our study shows that pembrolizumab treatment resulted in a successful remission of metastatic melanoma in a HIV-1-infected individual, and concomitantly enhanced the HIV-1 specific-CD8⁺ T-cell response in terms of activation and function. Nevertheless, this was not sufficient to consistently reduce the size of the viral reservoir, questioning the stand-alone effect of ICIs in terms of “shock and kill” HIV-1 curative strategies.

Supplementary Materials: The following are available online at <http://www.mdpi.com/2077-0383/8/12/2089/s1>, Figure S1: Immunophenotype of the HIV-1 specific-CD8⁺ T-cell response after 24 h of pembrolizumab administration from week 18 (w18+1).

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References

1. Ohaegbulam, K.C.; Assal, A.; Lazar-Molnar, E.; Yao, Y.; Zang, X. Human cancer immunotherapy with antibodies to the PD-1 and PD-L1 pathway. *Trends. Mol. Med.* **2015**, *21*, 24–33. [[CrossRef](#)]
2. Hamid, O.; Robert, C.; Daud, A.; Hodi, F.S.; Hwu, W.J.; Keir, R.; Wolchok, J.D.; Hersey, P.; Joseph, R.; Weber, J.S.; et al. Five-year survival outcomes for patients with advanced melanoma treated with pembrolizumab in KEYNOTE-001. *Ann. Oncol.* **2019**, *30*, 582–588. [[CrossRef](#)]

3. Trautmann, L.; Janbazian, L.; Chomont, N.; Said, E.A.; Gimmig, S.; Bessette, B.; Boulassel, M.-R.; Delwart, E.; Sepulveda, H.; Balderas, R.S.; et al. Upregulation of PD-1 expression on HIV-specific CD8+ T cells leads to reversible immune dysfunction. *Nat. Med.* **2006**, *12*, 1198. [[CrossRef](#)]
4. Day, C.L.; Kaufmann, D.E.; Kiepiela, P.; Brown, J.A.; Moodley, E.S.; Reddy, S.; Mackey, E.W.; Miller, J.D.; Leslie, A.J.; DePierres, C.; et al. PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease progression. *Nature* **2006**, *443*, 350. [[CrossRef](#)]
5. Hatano, H.; Jain, V.; Hunt, P.W.; Lee, T.-H.; Sinclair, E.; Do, T.D.; Hoh, R.; Martin, J.N.; Mccune, J.M.; Hecht, F.; et al. Cell-based measures of viral persistence are associated with immune activation and programmed cell death protein 1 (PD-1)-expressing CD4+ T cells. *J. Infect. Dis.* **2013**, *208*, 50–56. [[CrossRef](#)] [[PubMed](#)]
6. Fromentin, R.; Bakeman, W.; Lawani, M.B.; Khoury, G.; Hartogensis, W.; DaFonseca, S.; Killian, M.; Epling, L.; Hoh, R.; Sinclair, E.; et al. CD4+ T Cells Expressing PD-1, TIGIT and LAG-3 Contribute to HIV Persistence during ART. *PLOS Pathog.* **2016**, *12*, e1005761. [[CrossRef](#)] [[PubMed](#)]
7. DaFonseca, S.; Chomont, N.; El Far, M.; Boulassel, R.; Routy, J.; Sékaly, R. Purging the HIV-1 reservoir through the disruption of the PD-1 pathway. *J. Int. AIDS Soc.* **2010**, *13*, O15. [[CrossRef](#)]
8. Fromentin, R.; DaFonseca, S.; Costiniuk, C.T.; El-Far, M.; Procopio, F.A.; Hechy, F.M.; Hoh, R.; Deeks, S.G.; Hazuda, D.J.; Lewin, S.R.; et al. PD-1 blockade potentiates HIV latency reversal ex vivo in CD4+ T cells from ART-suppressed individuals. *Nat. Commun.* **2019**, *10*, 814. [[CrossRef](#)] [[PubMed](#)]
9. Deeks, S.G. Shock and kill. *Nature* **2012**, *487*, 439–440. [[CrossRef](#)] [[PubMed](#)]
10. Uldrick, T.S.; Gonçalves, P.H.; Abdul-Hay, M.; Claeys, A.J.; Emu, B.; Ernsto, M.S.; Fling, S.P.; Fong, L.; Kaiser, J.C.; Lacroix, A.M.; et al. Assessment of the Safety of Pembrolizumab in Patients With HIV and Advanced Cancer—A Phase 1 Study. *JAMA Oncol.* **2019**, *5*, 1332–1339. [[CrossRef](#)] [[PubMed](#)]
11. Cook, M.R.; Kim, C. Safety and E cacy of Immune Checkpoint Inhibitor Therapy in Patients with HIV Infection and Advanced-Stage Cancer. *JAMA Oncol.* **2019**, *5*, 1049–1054. [[CrossRef](#)] [[PubMed](#)]
12. Heppt, M.V.; Schlaak, M.; Eigentler, T.K.; Kähler, K.C.; Kiecker, F.; Loquai, C.; Meier, F.; Tomsitz, D.; Brenner, N.; Niesert, A.C.; et al. Checkpoint blockade for metastatic melanoma and Merkel cell carcinoma in HIV-positive patients. *Ann. Oncol.* **2017**, *28*, 3104–3106. [[CrossRef](#)] [[PubMed](#)]
13. Davar, D.; Wilson, M.; Pruckner, C.; Kirkwood, J.M. PD-1 Blockade in Advanced Melanoma in Patients with Hepatitis C and/or HIV. *Case Rep Oncol. Med.* **2015**, *2015*, 737389. [[CrossRef](#)] [[PubMed](#)]
14. Scully, E.P.; Rutishauser, R.L.; Simoneau, C.R.; Delagrèverie, H.; Euler, Z.; Thanh, C.; Li, J.Z.; Hartig, H.; Bakkour, S.; Busch, M.; et al. Inconsistent HIV reservoir dynamics and immune responses following anti-PD-1 therapy in cancer patients with HIV infection. *Ann. Oncol.* **2018**, *29*, 2141–2142. [[CrossRef](#)] [[PubMed](#)]
15. Le Gar, G.; Samri, A.; Lambert-Niclot, S.; Even, S.; Lavalé, A.; Cadranet, J.; Spano, J.P.; Autran, B.; Marcelin, A.G.; Guihot, A. Transient HIV-specific T cells increase and inflammation in an HIV-infected patient treated with nivolumab. *AIDS* **2017**, *31*, 1048–1051. [[CrossRef](#)] [[PubMed](#)]
16. Gay, C.L.; Bosch, R.J.; Ritz, J.; Hataye, J.M.; Aga, E.; Tressler, R.L.; Mason, S.W.; Hwang, C.K.; Grasela, D.M.; Ray, N.; et al. Clinical Trial of the Anti-PD-L1 Antibody BMS-936559 in HIV-1 Infected Participants on Suppressive Antiretroviral Therapy. *J. Infect. Dis.* **2017**, *215*, 1725–1733. [[CrossRef](#)] [[PubMed](#)]
17. Wightman, F.; Solomon, A.; Kumar, S.S.; Urriola, N.; Gallagher, K.; Hiener, B.; Palmer, S.; Mcneil, C.; Garsia, R.; Lewin, S.R. E ect of ipilimumab on the HIV reservoir in an HIV-infected individual with metastatic melanoma. *AIDS* **2015**, *29*, 504. [[CrossRef](#)] [[PubMed](#)]
18. Guihot, A.; Marcelin, A.-G.; Massiani, M.-A.; Samri, A.; Soulié, C.; Autran, B.; Spano, J.P. Drastic decrease of the HIV reservoir in a patient treated with nivolumab for lung cancer. *Ann. Oncol.* **2018**, *29*, 517–518. [[CrossRef](#)]
19. Breton, G.; Chomont, N.; Takata, H.; Fromentin, R.; Ahlers, J.; Filali-Mouhim, A.; Riou, C.; Boulassel, M.R.; Routy, J.P.; Yassine-Diab, B.; et al. Programmed death-1 is a marker for abnormal distribution of naive/memory T cell subsets in HIV-1 infection. *J. Immunol.* **2013**, *191*, 2194–2204. [[CrossRef](#)]
20. Roederer, M.; Nozzi, J.L.; Nason, M.C. SPICE: Exploration and analysis of post-cytometric complex multivariate datasets. *Cytom. Part A* **2011**, *79*, 167–174. [[CrossRef](#)]
21. Martínez-Bonet, M.; Puertas, M.C.; Fortuny, C.; Ouchi, D.; Mellado, M.J.; Rojo, P.; Noguera-Julian, A.; Muñoz-Fernández, M.A.; Martínez-Picado, J. Establishment and Replenishment of the Viral Reservoir in Perinatally HIV-1-infected Children Initiating Very Early Antiretroviral Therapy. *Clin. Infect. Dis.* **2015**, *61*, 1169–1178. [[CrossRef](#)] [[PubMed](#)]

22. Puertas, M.C.; Salgado, M.; Morón-López, S.; Ouchi, D.; Muñoz-Moreno, J.A.; Moltó, J.; Clotet, B.; Martínez-Picado, J. Effect of lithium on HIV-1 expression and proviral reservoir size in the CD4+ T cells of antiretroviral therapy suppressed patients. *AIDS* **2014**, *28*, 2157–2159. [[CrossRef](#)] [[PubMed](#)]
23. Amendola, A.; Bloisi, M.; Marsella, P.; Sabatini, R.; Bibbò, A.; Angeletti, C.; Capobianchi, M.R. Standardization and performance evaluation of 'modified' and 'ultrasensitive' versions of the Abbott RealTime HIV-1 assay, adapted to quantify minimal residual viremia. *J. Clin. Virol.* **2011**, *52*, 17–22. [[CrossRef](#)] [[PubMed](#)]
24. Ho, H.N.; Hultin, L.E.; Mitsuyasu, R.T.; Matud, J.L.; Hausner, M.A.; Bockstoe, D.; Chou, C.C.; O'Rourke, S.; Taylor, J.M.; Giorgi, J.V. Circulating HIV-specific CD8+ cytotoxic T cells express CD38 and HLA-DR antigens. *J. Immunol.* **1993**, *150*, 3070–3079.
25. Kestens, L.; Vanham, G.; Gigase, P.; Young, G.; Hannet, I.; Vanlangendonck, F.; Hulstaert, F.; Bach, B. Expression of activation antigens, HLA-DR and CD38, on CD8 lymphocytes during HIV-1 infection. *AIDS* **1992**, *6*, 793–798. [[CrossRef](#)]
26. Ruiz, A.; Blanch-Lombarte, O.; Jimenez-Moyano, E.; Ouchi, D.; Mothe, B.; Peña, R.; Galvez, C.; Genescà, M.; Martínez-Picado, J.; Goulder, P. Antigen Production After Latency Reversal and Expression of Inhibitory Receptors in CD8+ T Cells Limit the Killing of HIV-1 Reactivated Cells. *Front. Immunol.* **2019**, *9*, 3162. [[CrossRef](#)]
27. Velu, V.; Titanji, K.; Zhu, B.; Husain, S.; Pladevega, A.; Lai, L.; Vanderford, T.H.; Chennareddi, L.; Silvestri, G.; Freeman, G.J.; et al. Enhancing SIV-specific immunity in vivo by PD-1 blockade. *Nature* **2009**, *458*, 206. [[CrossRef](#)]
28. Akdis, M.; Aab, A.; Altunbulakli, C.; Azkur, K.; Costa, R.A.; Cramer, R.; Duan, S.; Eiwegger, T.; Eljaszewicz, A.; Ferstl, R.; et al. Interleukins (from IL-1 to IL-38), interferons, transforming growth factor, and TNF- α : Receptors, functions, and roles in diseases. *J. Allergy Clin. Immunol.* **2016**, *138*, 984–1010. [[CrossRef](#)]
29. Lenardo, M.; Chan, F.K-M.; Hornung, F.; McFarland, H.; Siegel, R.; Wang, J.; Zheng, L. Mature T lymphocyte apoptosis—immune regulation in a dynamic and unpredictable antigenic environment. *Annu. Rev. Immunol.* **1999**, *17*, 221–253. [[CrossRef](#)]
30. Gaen, S.; Liu, K. Overview of interleukin-2 function, production and clinical applications. *Cytokine* **2004**, *28*, 109–123. [[CrossRef](#)]
31. Aust, S.; Felix, S.; Auer, K.; Bachmayr-Heyda, A.; Kenner, L.; Dekan, S.; Meier, S.M.; Gerner, C.; Grimm, C.; Pils, D. Absence of PD-L1 on tumor cells is associated with reduced MHC I expression and PD-L1 expression increases in recurrent serous ovarian cancer. *Sci. Rep.* **2017**, *7*, 42929. [[CrossRef](#)] [[PubMed](#)]
32. Uldrick, T.S.; Fling, S.; Scott, V. Pembrolizumab induces HIV latency reversal in HIV+ individuals on ART with cancer. In Proceedings of the Conference on Retroviruses and Opportunistic Infections (CROI), Seattle, WA, USA, 4–7 March 2019; p. 27. Available online: <https://www.croiconference.org/abstracts/search-abstracts/> (accessed on 26 November 2019).
33. Banga, R.; Procopio, F.A.; Noto, A.; Pollakis, G.; Cavassini, M.; Ohmiti, K.; Corpataux, J.M.; de Leval, L.; Pantaleo, G.; Perreau, M. PD-1+ and follicular helper T cells are responsible for persistent HIV-1 transcription in treated aviremic individuals. *Nat. Med.* **2016**, *22*, 754. [[CrossRef](#)] [[PubMed](#)]
34. González-Cao, M.; Moran, T.; Dalmau, J.; Garcia-Corbacho, J.; Bernabé, R.; Juan, O.; de Castro, J.; Blanco, R.; Meyerhand, A.; Blanco, J.; et al. Phase II study of durvalumab (MEDI4736) in cancer patients HIV-1-infected. *Clin. Oncol.* **2019**, *37*, 2501. [[CrossRef](#)]

